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CheKine™ Micro Nitric Oxide (NO) Assay Kit

Cat #: KTB1400 Size: 48 T/48 S 96 T/96 S

[<u>=</u> Q	Micro Nitric Oxide (NO) Assay Kit				
REF	Cat #: KTB1400	LOT	Lot #: Refer to product label		
	Applicable samples: Animal and Plant Tissues, Cells, Plasma, Serum, Urine (and other Biological Fluids)				
Å	Storage: Storage at -20°C for 12 months, protected from light				

Assay Principle

Nitric oxide (NO) is a reactive radical that plays an important role in many key physiological functions such as neurotransmission, immune response and apoptosis. NO is synthesized from L-arginine by NO synthase (NOS). It has been identified as an endothelial derived relaxation factor (EDRF) and antiplatelet substance. It serves as a neurotransmitter derived from a neutrophil and a cytotoxic substance from an activated macrophage. Although NO's molecular action in the biological system is very versatile, the most important role of NO is the activation of guanylate cyclase. CheKine™ Micro Nitric Oxide (NO) Assay Kit is designed to accurately measure NO production following reduction of nitrate to nitrite by using improved griess method. The Griess assay's mechanism is summarized as the azo coupling between diazonium species, which are produced from sulfanilamide with NO²- and N-(1-naphthyl) ethylenediamine dihydrochloride, resulting in a colorimetric (540 nm) product proportional to the NO metabolite present.

Materials Supplied and Storage Conditions

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Kit components	48 T	96 T	Storage conditions	
NaNO ₂ Standard (1 M)	1 mL	1 mL	-20°C, protected from light	
Griess Reagent	4 mL	8 mL	4°C, protected from light	
Griess Reagent	4 mL 8 mL		4°C, protected from light	
VCl ₃ Reagent	7.5 mL	15 mL	4°C, protected from light	
ZnSO ₄	0.6 mL	1.2 mL	4°C, protected from light	

Note: Before formal testing, it is recommended to select 2-3 samples with large expected differences for pre-experiment.

Materials Required but Not Supplied

- · Microplate reader or visible spectrophotometer capable of measuring absorbance at 540 nm
- Incubator
- 96-well plate or microglass cuvette, precision pipettes, disposable pipette tips



- Deionized water, PBS (pH 7.4)
- Dounce homogenizer (for tissues samples)

Reagent Preparation

NaNO₂ Standard (1 M): Equilibrate to room temperature and protect from light during the assay. Store aliquots at -20°C, and protected from light.

Griess Reagent I: Ready to use as supplied. Equilibrate to room temperature and protected from light during the assay. Store at 4°C, and protected from light.

Griess Reagent II: Ready to use as supplied. Equilibrate to room temperature and protected from light during the assay. Store at 4°C, and protected from light.

VCI₃ Reagent: Ready to use as supplied. Equilibrate to room temperature and protected from light during the assay. Store at 4°C, and protected from light.

ZnSO₄: Ready to use as supplied. Equilibrate to room temperature and protected from light during the assay. Store at 4°C, and protected from light.

Standard preparation: Prepare 10 mM of NaNO₂ Standard stock solution | by diluting 10 μ L NaNO₂ Standard (1 M) into 990 μ L PBS (pH 7.4). Prepare 100 μ M of NaNO₂ Standard stock solution || by diluting 10 μ L 10 mM of NaNO₂ Standard into 990 μ L PBS (pH 7.4). Using 100 μ M standard stock solution || , prepare standard curve dilution as described in the table:

Num.	100 μM Standard stock solution II (μL)	PBS (pH 7.4) (μL)	Concentration (µM)
Std.1	200	0	100
Std.2	100	100	50
Std.3	40	160	20
Std.4	20	180	10
Std.5	10	190	5
Std.6	4	196	2
Std.7	2	198	1
Blank	0	200	0

Note: Always prepare fresh standards per use. Diluted standard solution is unstable and must be used within 4 h.

Sample Preparation

- 1. Animal and Plant Tissues or Cell samples: Weigh 0.1 g tissue or collect 5x10⁶ cells add 1 mL PBS (pH 7.4) and homogenize. Centrifuge at 14,000 rpm for 10 min at 4°C. Use supernatant for NO assay.
- 2. Plasma, Serum and Urine (and other Biological Fluids): Tested directly by adding samples to the microplate wells. Samples that need to conduct deproteinization include serum, plasma, whole blood, cell culture media containing FBS, tissue or cell lysates. urine and saliva do not need deproteinization.

Deproteination: Mix 150 μ L sample with 8 μ L ZnSO₄ in 1.5 mL tubes. Vortex and then centrifuge at 14,000 rpm for 10 min at 4°C. Transfer 100 μ L of the clear supernatant to a clean tube.

Note: We recommend that you use fresh samples. If not assayed immediately, samples can be stored at -80°C for one month. If samples need to be deproteinated, 150 μ L of each standard should be prepared and also be treated with ZnSO₄ to eliminate the need for a dilution factor. Antioxidants and nucleophiles (e.g. β -mercaptoethanol, glutathione, dithiothreitol and cysteine) may interfere with this assay. Avoid using these compounds during sample preparation.

Assay Procedure

1. Preheat the microplate reader or visible spectrophotometer for more than 30 min, and adjust the wavelength to 540 nm, Visible



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spectrophotometer was returned to zero with deionized water.

- 2. Working Reagent preparation: For each well of reaction, prepare 204 μL Working Reagent by mixing 104 μL VCl₃ Reagent, 50 μL Griess Reagent | and 50 μL Griess Reagent | . Working Reagent is freshly prepared.
- 3. Add 100 µL diluted Standards and Sample to separately labeled EP tubes (We recommend that Samples be measured in at least duplicate). Then add 200 µL Working Reagent to each Sample and Standard tube, then mix well.
- 4. Incubate the reaction for 30 min at 37°C.
- 5. Briefly centrifuge the reaction tubes to pellet any condensation and transfer 250 μ L of each reaction to separate wells in a 96 -well plate or microglass cuvette. Read A at 540 nm. and recorded as A_{Blank} , $A_{Standard}$ and A_{Sample} , respectively. Subtract A_{Blank} from $A_{Standard}$ and A_{Sample} , that is, $\Delta A_{Standard}$ = $A_{Standard}$ ΔA_{Sample} Δ

Note: In order to guarantee the accuracy of experimental results, need to do a pre-experiment with 2-3 samples. If the A_{Sample} is higher than 1.0, please further dilute the sample with PBS (pH 7.4). Pay attention to multiply by the dilution factor when calculating the result. Increase the sample size if the ΔA_{Sample} value is below 0.005.

Data Analysis

1. Calculation of NO content

With the concentration of the Standard Solution as the x-axis and the $\Delta A_{Standard}$ as the y-axis, draw the standard curve. Substitute the ΔA_{Sample} into the equation to obtain the x value (μM).

- (1) Calculated by weight of samples
- NO content (nmol/g)= $x \times V_{Sample} \div (W \times V_{Sample} \div V_{total\ sample}) \times n = x \div W \times n$
- (2) Calculated by protein concentration
- NO content (nmol/mg prot)=xxVsample÷(VTotal samplexCpr)xn=x+Cprxn
- (3) Calculated by volume of Liquid samples
- NO content (nmol/ml)=xxVTotal liquid ÷VLiquid sample xn=xx1.05xn (Deproteination treated sample)
- NO content (nmol/ml)=xxV_{Liquid sample}÷V_{Liquid sample}×n=xxn (non- deproteination treated sample)
- (4) Calculated by number of cells
- NO content (nmoL/10⁴ cells)=xxV_{Sample}÷(NxV_{Sample}÷V _{Sample})xn =x÷Nxn

Where: nmol: 1 μ M=1 nmol/mL; V_{Sample}: added sample volume, 0.1 mL; V_{Total liquid}: total volume of liquid sample and ZnSO₄, 0.158 mL; V_{Liquid sample}: liquid sample volume, 0.15 mL; n: the sample dilution factor; Cpr: sample protein concentration, mg/mL; W: weight of sample, g; V_{Total sample}: total sample extraction volume, 1 mL; N: number of cells, 1×10⁴.

Typical Data

Typical standard curve-data provided for demonstration purposes only. A new standard curve must be generated for each assay performed.

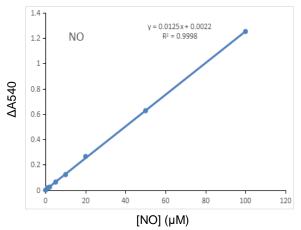


Figure 1. Standard Curve of NO in 96-well plate assay.



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Recommended Products

Catalog No.	Product Name
KTB1010	CheKine™ Micro Coenzyme II NADP(H) Assay Kit
KTB1020	CheKine™ Micro Coenzyme ∣ NAD(H) Assay Kit
KTB1030	CheKine™ Micro Superoxide Dismutases (SOD) Activity Assay Kit
KTB1070	CheKine™ Micro Xanthine Oxidase Activity Assay Kit

Disclaimer

The reagent is only used in the field of scientific research, not suitable for clinical diagnosis or other purposes. For your safety and health, please wear a lab coat and disposable gloves.

