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Anti-DDDDK Tag Mouse Monoclonal Antibody (1B10), Magnetic Beads

Cat #: A02010MGB Size: 100μl /400μl /2ml

Product Information

	Product Name: Anti-DDDDK Tag Mouse Monoclonal Antibody (1B10), Magnetic Beads		
	Applications: IP		**Isotype:** Mouse IgG1
REF	**Catalog Number:** A02010MGB	LOT	**Lot Number:** Refer to product label
	**Storage:** Store at 4°C. Avoid freeze-thaw or centrifugation.		**Note:** Contain sodium azide.
	Formulation: Over 1 mg of Antibody coupled to 1 ml of packed Magnetic beads.		

Product Description: Anti-DDDDK Tag Magnetic Beads are prepared by covalently coupling Anti-DDDDK Tag Mouse Monoclonal Antibody to crosslinked Magnetic Beads, useful for detection and capture of fusion proteins containing a DDDDK peptide sequence by commonly used immunoprecipitation procedures. The coupling technique is optimized to give a high binding capacity for DDDDK-tag protein.

Storage Buffer: Suspended in PBS, pH 7.4, containing 0.02% Sodium Azide as preservative.

Storage instructions: Stable for one year at 4°C from date of shipment. Avoid freeze-thaw or centrifugation.

Shipping: Gel pack with blue ice.

Note: The product listed herein is for research use only and is not intended for use in human or clinical diagnosis. Suggested applications of our products are not recommendations to use our products in violation of any patent or as a license. We cannot be responsible for patent infringements or other violations that may occur with the use of this product.

Suggested Procedure

Magnetic Beads Preparation (Repeat for three times)

1. Transfer the appropriate amount of Magnetic beads (Suggested volume: 10-20 μL Magnetic beads for each sample) into a 1.5 mL microfuge tube and place it on the Magnetic separator for 10s.
2. Remove the storage solution.
3. Resuspend the Magnetic beads with 1 mL ice-cold 1×TBS.
4. Place it on the Magnetic separator for 10 s and remove the supernatant.

Note: For multiple samples, it is recommended to prepare the Magnetic beads first, and then aliquot into each microfuge tube.

Sample preparation

1. Collect 2×10^6 cells and wash with PBS for three times.
2. Resuspend cells in 400 μL ice-cold lysis buffer [50 mM Tris (pH 7.5), 150 mM NaCl, 0.05% NP-40], then sonicate briefly (up to 10s).
3. Centrifugate 12000*g* for 5 minutes at 4°C and collect the supernatant.

Binding protein

1. Add 400 μL cell lysate to Magnetic beads.
2. Mix thoroughly and incubate at 4°C for 1-2 h.

Note: During the binding process, the Magnetic beads may be clustered or flake. This phenomenon is normal and will not affect the experimental results.

Wash (Repeat 3-5 times until OD280 of the supernatant is lesser than 0.05)

1. Place the tube on the Magnetic separator for 10s to collect the mixture.
2. Resuspend the mixture with 1 mL ice-cold 1×TBS and incubate it at 4ºC for 1-3 minutes.
3. Place it on the Magnetic separator for 10 s and remove the supernatant. (Or save the supernatant for further analysis)

Note: Avoid losing Magnetic Beads during wash step.

Elution

For SDS-PAGE detection -- Add 50 μL 1×protein sample buffer to the above obtained precipitate, and then boil for 5 min. Cool to room temperature and place in on the Magnetic separator for 10s.

For other assay -- Add 2 volumes (vs beads volume) elution buffer [0.1 M -0.2 M Glycine pH 2.5-3.1 (or 0.1 M citric acid, pH 2.5-3.1 or 2.5% Acetic Acid)] to the above obtained precipitate, and incubate at least 2 minutes to collect elution fraction. (Repeat for three times).